

Synthesis of C₂-Symmetric Guanidino-Sugars as Potent Inhibitors of Glycosidases[†]

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Abstract—A series of enantiomerically pure C_2 -Symmetric guanidino-sugars was synthesized from D-mannitol. The first method described involves direct opening of a bis-epoxide by guanidine, whereas the second one deals with a mercury-catalyzed transformation of a cyclic thiourea into a N,N',N''-trisubstituted guanidine as a key step. The biological activity of these compounds towards several glycosidases has been evaluated. One of them (5) was found to selectively inhibit α -L-fucosidase of bovin kidney (2.8 μ M). © 2000 Elsevier Science Ltd. All rights reserved.

Introduction

Glycosidases are enzymes which are responsible for the hydrolysis of the glycosyl bond; they play a key role in glycoprotein trimming, catabolism of glycoconjugates and degradation of polysaccharides. As a consequence, specific reversible inhibitors of those enzymes can have major therapeutic utility in the treatment of diabetes, 1 cancer² and viral infections.³

The strategy commonly encountered to design such inhibitors is to synthesize compounds that are able to mimick the oxocarbenium-type transition state,⁴ which is supposed to arise during glycosyl bond hydrolysis. In most cases, azasugars^{5–13} or amidine^{14–21} derivatives have been chosen in order to resemble the transient oxonium. More recently, guanidino-sugars^{22–24} have been proposed as a new class of glycosyl cation mimics.

Moreover, in the aim of synthesizing compounds capable of interacting with glycosidases, it has been shown that the aglycon part of the glycoside also plays an important role in the interaction. In particular, pseudo-disaccharides^{25–29} or pseudo-sugars containing an alkyl aglycone part³⁰ deserve more consideration.

Keywords: D-mannitol; guanidine; α -L-fucosidase inhibitor; mercuric chloride; thiourea.

Here we report full results on both the synthesis of C_2 -Symmetric guanidino-sugars 1, 2 and 3, 4^{31} from D-mannitol (Scheme 1), and the introduction on compound 4 of a *n*-butyl or glucopyranosyl group as an aglycon part (respectively, 5 and 6). Results on the biological evaluation of those compounds as potent inhibitors of glycosidases will be presented.

Compounds 1, 2 and 3, 4 are respectively analogues of the azepanes 7, 8 and of the pyrrolidines 9, 10, we precedently described and that inhibit glycosidases in low micromolar range.³² Two retrosynthetic pathways (Scheme 2) have been imagined as a solution to this challenging synthesis. The first one involves the heterocyclization of bis-electrophiles derived from D-mannitol by guanidine. The use of bis-epoxydes will lead to cyclic guanidine 1 or 2, while an activation at positions 2 and 5 of D-mannitol will lead to 3 or 4. The second one involves the hetero-cyclization of a diamine as a bisnucleophile.

Results and Discussion

The commercially available guanidine in salt form (hydrochloride, carbonate, nitrate or sulfate) has already been used as a nucleophile in reactions with α -diketones, 33 α , β -ethylenic esters 34 and alkyl halides, 35 and more recently in the opening of epoxides. For example, guanidine reacts with benzene trioxide 36 to lead cyclic guanidine, and with the 1,2:5,6-dianhydro-3,4-di-O-

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[†]Warmly dedicated to Professeur Pierre Sinaÿ on the occasion of his 62nd birthday.

Scheme 1.

Scheme 2.

benzyl-L-iditol³⁷ to give the corresponding guanidinomethyl tetrahydrofuran by subsequent *O*-cyclization. In the latter case, considering that the protective groups play a decisive role in the selectivity of ring opening of the bis-epoxide, we chose to protect the 3,4-diol as an acetonide to prevent *O*-cyclization; the corresponding bis-epoxides 11 and 12 (Scheme 3) being easily prepared from D-mannitol.^{38,39}

Thus, treatment of 1,2:5,6-dianhydro-3,4-*O*-methylethylidene-D-mannitol **11** (or L-iditol **12**) by 1 equivalent of guanidine (generated by ethanolic elution of its hydrochloride salt on Amberlite[®] IRA 400 (OH⁻)⁴⁰) in refluxing ethanol afforded the corresponding C₂-Symmetric cyclic guanidine **13** (or **14**) in 97% yield. Removal of the acetonide group was easily achieved with hydrogen chloride in methanol or with aqueous trifluoroacetic acid.

Scheme 3. (a) Guanidine, EtOH Δ , 97%; (b) HClg, MeOH, 70%; (c) TFA, H₂O, 70%.

According to the same strategy, the synthesis of cyclic guanidines 3–6 supposed the nucleophilic substitution of alcohols in positions 2 and 5 of D-mannitol or L-iditol, after protection of all the other alcohol functions. Thus, remaining secondary alcohols could be activated as mesylates or by the Mitsunobu reaction in order to react with guanidine. 41,42 Both approaches have been tried on different protected derivatives 15, 16, 17, or 18 (Scheme 4), offering different spatial restraints. Diols 16⁴³ and 17^{44,45} were prepared as already described, whereas the diol 15 was easily obtained from the commercially available 3,4-O-methylethylidene-D-mannitol 19 by selective benzylation of both primary hydroxyl groups through stannylidene activation with Bu₂SnO^{46,47} (Scheme 5), and subsequent treatment with benzyl bromide (86%). Lastly, the diol 18 was prepared from the dicarbonate 20^{48,49} by protection of the alcohol functions in positions 3 and 4 as trioxepane⁵⁰ according to a method developed by Beck.⁵¹ Reaction of dicarbonate 20 with paraformaldehyde and BF₃.Et₂O led to an unseparable

Scheme 4.

mixture of **21** and **22** (71% yield in 3:1 ratio according to ¹H NMR). Hydrolysis of carbonates in presence of pyridine yielded a mixture of the corresponding tetrols **23** and **24**. Final protection of both primary alcohols with Bu₂SnO and benzyl bromide, as described above, afforded the corresponding mixture of **18** and **25** which could be separated by flash chromatography (30 and 8% overall yield from **20**, respectively).

On one hand, the dimesylate **26**, **27** or **28**, issued, respectively, from diol **15**, **16** or **17** after treatment with methanesulfonyl chloride, failed to react with different sources of guanidine (free guanidine, acetyl-guanidine, N,N'-di-benzyloxy-carbonyl-guanidine), and on the other hand, Mitsunobu reaction involving N,N'-di-benzyloxy-carbonyl-guanidine on diol **16** or **18** led to the corresponding tetrahydrofuran **29** or **30** (Scheme 6). In the absence of the guanidine derivative, the yield of **29** or **30** could even be improved up to 91 or 68%, respectively.

Thus, this first strategy involving the *N,N'*-dialkylation of guanidine revealed to only be efficient with a 1,6-bis activated derivative of D-mannitol. In order to obtain the cyclic guanidines **3–6** we turned our attention to the second strategy dealing with the cyclization of a 2,5-diamine derived from D-mannitol or L-iditol into thiourea and subsequent transformation into guanidine.

Synthesis of the readily available cyclic thiourea 33 via the L-ido-diamine 32 is depicted in Scheme 7. Nucleophilic substitution of the 2,5-dimesylate 26 by sodium azide afforded the diazido derivative 31 (57% overall yield from 15) with inversion of configuration at C_2 and C_5 , then subsequent heterogeneous catalytic reduction led to the corresponding diamine 32 (87%). Cyclization of 32 into thiourea 33 smoothly occurred by treatment with carbon disulfide^{52,53} in pyridine (86%).

In order to transform the thiourea 33 into guanidine, the well-known method of thiourea alkylation into inter-

16: P = Bn **29**: P = Bn

18: $P = -(CH_2OCH_2)$ - **30**: $P = -(CH_2OCH_2)$ -

Scheme 6.

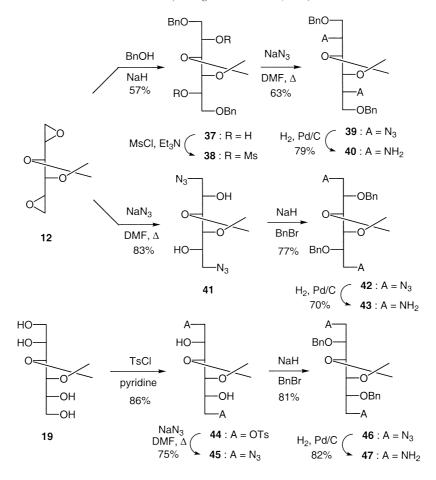
Scheme 7.

Scheme 8. (a) MeOTs, MeOH Δ, 88%; (b) BnNH₂, HgCl₂, Et₃N, CH₂Cl₂, 91%; (c) Na, NH₃, 74%; (d) TFA, H₂O, *quant*.

mediary methylisothiourea 34^{54–57} was first tried (Scheme 8). Although methylation at the exocyclic sulfur atom of 33 with methyl para-toluene-sulfonate⁵⁸ in refluxing methanol was easily achieved (88%), all attempts to substitute the thiomethyl group either with ammonia⁵⁹ or benzylamine⁶⁰ failed. We therefore turned to a more direct method using HgCl₂.61,62 Such an approach had already successfully been used on Ntert-butyloxycarbonyl thioureas in DMF but was reported to be inefficient on unactivated thioureas.⁶³ Indeed, this method involving the thiourea 33 in the presence of benzylamine and CH₂Cl₂ as solvent,⁶⁴ revealed to efficiently afford the N-benzyl cyclic guanidine 35 (91%). The last step was the deprotection of cyclic guanidine 35. Concerning reduction of benzyl ethers and N-benzyl guanidine, different attempts involving hydrogeneous catalytic reduction failed, while the use of sodium in liquid ammonia was successful. After 6h, the reaction was quenched by addition of solid ammonium chloride. Then, an extraction with absolute ethanol was followed by discarding ammonium chloride in excess by an ethanolic elution on Amberlite® IRA 400 (OH⁻). Final hydrolysis of the acetonide group was achieved in aqueous trifluoroacetic acid to yield the guanidino-sugar 4.

This strategy could also be applied to the synthesis of cyclic guanidines 3, 2 and 1 from corresponding diamines 40, 43 and 47 (Scheme 9). Interestingly, this second way of synthesizing cyclic guanidines 1 and 2 offered a justification for their structures. In that purpose, D-manno diamine 40 and L-ido diamine 43 were obtained from L-ido bis-epoxide 12. On one hand, opening of bis-epoxide 12 with sodium benzylate cleanly occurred to give the 1,6-di-O-benzyl derivative 37 (57%) which was then routinely converted into the 2,5-diamine 40 as described above for diamine 32 from diol 15. On the other hand, opening of bis-epoxide 12 by sodium azide led to the diol 41 (83%) which was protected as dibenzyl ether 42, by successive treatment with sodium hydride and benzyl bromide (77%). Finally, heterogeneous catalytic reduction in ethanol led to the diamine 43 (70%). The last diamine 47 was obtained from tetrol 19 as follows: selective tosylation of both primary hydroxyl groups with para-toluenesulfonyl chloride in pyridine³⁹ (86%), then nucleophilic substitution with sodium azide (75%) and heterogeneous catalytic reduction (82%). Cyclization of diamines 40, 43 and 47 into thioureas 48, 51 and 53 and subsequent transformations into the corresponding guanidino-sugars are summarized in Table 1.

Furthermore, this strategy which involves transformation of a cyclic thiourea into guanidine with benzylamine could be extended to other primary amines such as *n*-butylamine or methyl 6-amino-6-deoxy-2,3,4-tri-*O*-benzyl-D-gluco-pyranoside⁶⁵ (Table 2). Thus, *n*-butylamine treatment of thiourea 33 in CH₂Cl₂ in the presence of mercuric chloride at 20 °C led to the cyclic *N*-butyl guanidine 55 (65%), while reaction with the 6-amino-6-deoxy derivative of D-glucose in 1,2-dichloroethane at 80°C afforded the protected pseudo-disaccharide 57 (86%). By sodium reduction and subsequent acidic hydrolysis, as above, 55 and 57 yielded the corresponding deprotected guanidino-sugars 5 and 6.



Scheme 9.

Biological activity of synthesized guanidino-sugars towards different glycosidases (α - and β -D-glucosidases, α -D-mannosidase and α -L-fucosidase) was evaluated (Table 3). The results show that guanidino-sugars are weak inhibitors of glycosidases. For example, 3 is a less potent inhibitor of α - and β -D-glucosidases than its parent azasugar 9 (DMDP). The best results observed concerned the *N*-butyl and the *N*-glucosyl derivatives 5 and 6 which competitively inhibit α -L-fucosidase, K_i = 2.8 and 500 μM, respectively. Selective strong inhibition of α -L-fucosidase by compound 5, and more weakly by compound 6 confirms that the aglycon part plays a critical role in recognition. In particular, an alkyl aglycon moiety may be decisive in order to inhibit α -L-fucosidase.

Experimental⁶⁶

General methods

¹H NMR (250 MHz) and ¹³C NMR (63 MHz) spectra were recorded in CDCl₃ (unless otherwise indicated) on a Bruker ARX 250 spectrometer. Optical rotations were measured on a Perkin–Elmer 241C polarimeter with sodium lamp (589 nm) at 20 °C. IR spectra were recorded on a Perkin–Elmer 783 spectrophotometer and wave-numbers of characteristic absorption bands are given in cm⁻¹. Elemental analyses were performed by the 'Service Régional de Microanalyse de l'Université Pierre et Marie Curie',

Paris, France. Mass spectra were performed by the 'Service de Spectrométrie de Masse de l'Ecole Normale Supérieure', Paris, France. All reactions were monitored by thin-layer chromatography with Merck 60F 254 precoated silica (0.2 mM) on glass. Flash chromatography was performed with Merck Kieselgel 60H (5–40 μM). Prior to use, THF and Et₂O were distilled from benzophenone-sodium, and CH₂Cl₂ from CaH₂. Spectroscopic (¹H and ¹³C NMR, MS) and/or analytical data were obtained using chromatographically homogeneous samples.

(5*R*,6*R*,7*R*,8*R*)-5,6,7,8-Tetrahydroxy-1,3-diazonan-2-iminium chloride (1). A solution of the guanidine 13 (50 mg, 21 μmol) in gazeous hydrogen chloride saturated methanol (1 mL) was heated at 60 °C during 1 h. After cooling to 20 °C, the solid was filtered and washed with ethanol to yield the guanidinium chloride 1 (35 mg, 70%) as a white hygroscopic solid. [α] +5 (c 0.8, H₂O); ¹H NMR (D₂O): 3.35–3.50 (m, 2H, H_{5,8}), 3.60–3.75 (m, 2H, H_{6,7}), 3.75–4.05 (m, 4H, H_{4,9}); ¹³C NMR (D₂O): 47.9 (C_{4,9}), 71.6, 72.2 (C_{5,6,7,8}), 160.4 (C₂); HRMS (FAB⁺) for C₇H₁₆O₄N₃: (MH⁺) calcd 206.1141, found: 206,1118.

(5S,6R,7R,8S)-5,6,7,8-Tetrahydroxy-1,3-diazonan-2-iminium trifluoroacetate (2). A solution of the guanidine 14 (180 mg, 76 μmol) in 10% aq trifluoroacetic acid (4 mL) was heated at 60 °C during 1 h. After concentration in vacuo, the resulting oil was triturated with diethylether to leave a white solid collected by filtration and washed with

Table 1.

Diamine	Thiourea	Protected guanidine	Guanidino-sugar
BnO H ₂ N O NH ₂ OBn 40	BnO NH NH S 48 (81%)	PO NH NP OP NP 49: P = Bn (78%)	HO OH HN ⊕ NH CF ₃ CO ₂ NH ₂ 3 (100%)
40	40 (01 /0)	50: $P = H (75\%)$	3 (100 %)
H ₂ N OBn O NH ₂	BnO,,,,,, OBn HN NH S 51 (100%)	PO NP 52: P = Bn (94%) 14: P = H (80%)	HO OH HO OH CF ₃ CO ₂ NH ₂ 2 (70%)
H ₂ N BnO O OBn NH ₂	BnO NH S 100%)	14. P - H (80%) PO NH NP 54: P = Bn (94%) 13: P = H (77%)	HO OH HN + NH CF ₃ CO ₂ NH ₂

diethylether to yield the guanidinium trifluoroacetate **2** (177 mg, 70%) as a white hygroscopic solid. [α] + 19 (c 0.3, H₂O); ¹H NMR (D₂O): 3.25–3.40 (m, 2H, H_{6,7}), 3.45–4.10 (m, 6H, H_{4,5,8,9}); ¹³C NMR (D₂O): 54.7 (C_{4,9}), 74.2, 76.1 (C_{5,6,7,8}), 159.6 (C₂).

(4*R*,5*R*,6*R*,7*R*)-5,6-Dihydroxy-4,7-dihydroxy methyl-1, 3-diazepan-2-iminium trifluoroacetate (3). The guanidinium trifluoroacetate 3 (40 mg) was quantitatively obtained as a whitish hygroscopic solid from the guanidine **50** on a 0.12 mmol scale, according to the procedure described above for the guanidino-sugar **2**. [α] +8 (*c* 1.3, H₂O); ¹H NMR (D₂O): 3.40–3.50 (m, 2H, H_{4,7}), 3.60–3.70 (m, 2H, H_{5,6}), 3.93 (dd, 2H, $J_{4,CHOH}$ = 6.4, J_{CH_2OH} = 12.0 Hz, CH₂OH), 4.06 (dd, 2H, $J_{4,CH'OH}$ = 3.6, J_{CH_2OH} = 12.0 Hz, CH₂OH); ¹³C NMR (D₂O): 60.7 (C_{4,7}), 63.2 (CH₂OH), 75.3 (C_{5,6}); HRMS (FAB⁺) for C₇H₁₆ N₃O₄: (MH⁺) calcd 206.1141, found 206.1095.

(4*S*,5*R*,6*R*,7*S*)-5,6-Dihydroxy-4,7-dihydroxy methyl-1,3-diazepan - 2 - iminium trifluoroacetate (4). The guanidinium trifluoroacetate 4 (27 mg) was quantitatively obtained as a whitish hygroscopic solid from the guanidine 36 on a 0.08 mmol scale, according to the procedure described above for the guanidino-sugar 2. $[\alpha] + 12$ (c

1.0, H₂O); ¹H NMR (D₂O): 3.65–4.10 (m, 8H, H_{4,5,6,7}, CH₂OH); ¹³C NMR (D₂O): 59.6 (C_{4,7}), 63.9 (CH₂OH), 72.3 (C_{5,6}), 160.6 (C₂); HRMS (FAB⁺) for C₇H₁₆N₃O₄: (MH⁺) calcd 206.1141, found 206.1165.

(4*S*,5*R*,6*R*,7*S*)-5,6-Dihydroxy-4,7-dihydroxy methyl-1,3-diazepan-2-(*N*-butyl)-iminium trifluoroacetate (5). The guanidino-sugar 5 (120 mg, 96%) was obtained as a whitish hygroscopic solid from the guanidine 58 on a 0.33 mmol scale, according to the procedure described above for the guanidino-sugar 2. [α] +2 (c 1.0, MeOH); ¹H NMR (CD₃OD): 0.97 (t, 3H, $J_{3',4'}$ =7.3 Hz, $H_{4'}$), 1.37 (m, 2H, $H_{3'}$), 1.64 (m, 2H, $H_{2'}$), 3.65–4.00 (m, 8H, $H_{4,7,1'}$, CH₂OH), 4.17 (m, 2H, $H_{5,6}$); ¹³C NMR (CD₃OD): 15.8 (C_{4'}), 22.8 (C_{3'}), 33.7 (C_{2'}), 45.2 (C_{1'}), 59.4 (C_{4,7}), 65.8 (CH₂OH), 80.0 (C_{5,6}), 160.6 (C₂); MS (NH₃): 262 (MH⁺).

Methyl (4'S,5'R,6'R,7'S)-6-deoxy-D-gluco pyranoside-6-(4',7'-dihydroxy methyl-5',6'-dihydroxy-1',3'-diazepan-2'-yl)-iminium trifluoroacetate (6). The guanidino-sugar 6 (160 mg, 98%) was obtained as a whitish hygroscopic solid from the guanidine 60 on a 0.33 mmol scale according to the procedure described above for guanidino-sugar 2. $[\alpha] + 25$ (c 0.9, CH₃OH); ¹H NMR (CD₃OD): 3.35–3.43 (m, 11H, H_{4.7}, CH₂N, OCH₃, CH₂OH), 3.69–3.77

Table 2.

Reactants	Protected guanidine	Guanidino-sugars	
33 + <i>n</i> BuNH ₂ , HgCl ₂ ,CH ₂ Cl ₂ , 20 °C	PO HN NH	HO OH HN ⊕ NH CF ₃ CO ₂ HN	
33 + ,NH ₂	55: P = Bn (65%) 56: P = H (73%)	5 (96%)	
BnO BnO Me	PO HN NH	HO OH OH	
HgCl ₂ ,ClCH ₂ CH ₂ Cl, 80 °C	PO PO Me	CF ₃ CO ₂ NH HO HO HO OMe	
	57 : P = Bn (86%) 58 : P = H (79%)	6 (98%)	

Table 3.

Enzyme	Inhibitors (%) ^a					
	1	2	3	4	5	6
α-D-glucosidase ^b	24	36	14	37	41	14
β-D-glucosidase ^c	7	34	10	16	NI	6
α-D-mannosidase ^d	10	41	4	4	18	12
α-L-fucosidase ^e	21	31	12	32	94	96
					$2.8\mu M^f$	$500 \mu M^f$

^a% Inhibition determined at 1 mM concentration of inhibitor; NI for no inhibition.

(m, 6H, $H_{5,6,2',3',4',5'}$), 4.77 (m, 1H, $H_{1'}$); ¹³C NMR (CD₃ OD): 46.0 (CH₂N), 52.5, 58.7 ($C_{4,7}$, OCH₃), 66.0 (CH₂ OH), 71.8, 74.3, 74.9, 75.8, 77.1 ($C_{5,6,2',3',4',5'}$), 103.8 ($C_{1'}$); MS (FAB⁺): 382 (MH⁺).

(5*R*,6*R*,7*R*,8*R*)-5,6,7,8-Tetrahydroxy-6,7-*O*-methylethylidene-2-imino-1,3-diazonane (13)

From 1,2:5,6-dianhydro-3,4-*O*-methylethylidene-D-mannitol (11). A solution of guanidinium chloride (83 mg, 87 μmol, 0.95 equiv) in a minimum of 95% ethanol:water was neutralized on Amberlite[®] IRA 400 (OH⁻), then eluted with absolute ethanol. After concentration in vacuo, free guanidine was dissolved in absolute ethanol (1 mL), the bis-epoxide 11³⁹ (170 mg, 92 μmol) was added and the resulting mixture was heated under reflux during 1 h. After concentration in vacuo, water and

CH₂Cl₂ were added. The organic layer was washed with water and the combined aqueous layers were concentrated in vacuo to yield the guanidine **13** (208 mg, 97%) as an hygroscopic solid.

From (5R,6R,7R,8R)-5,8-dibenzyloxy-7,8-dihydroxy-7, 8-di-*O*-methylethylidene-2-benzylimino-1,3-diazonane (54). A solution of the benzylated guanidine 54 (50 mg, 97 µmol) in a minimum of anhydrous tetrahydrofuran was added into liquid ammonia (5 mL). After addition of sodium until persistance of a dark blue color, the reaction mixture was stirred under refluxing ammonia during 6h. Then, careful addition of solid ammonium chloride resulted in decoloration and was followed by ammonia evaporation. The resulting solid was washed with CH₂Cl₂ and extracted with absolute ethanol. The combined alcoholic layers were concentrated in vacuo, neutralized over Amberlite® IRA 400 (OH⁻), eluted with ethanol and concentrated in vacuo to yield the guanidine 13 (23 mg, 77%) as an hygroscopic solid. $[\alpha] + 24$ (c 1.0, MeOH); ¹H NMR (D₂O): 1.47 (s, 6H, CH₃), 3.38– 3.58 (m, 4H, H_{4,9}), 3.68–4.00 (m, 4H, H_{5.6,7.8}); ¹³C NMR (CD_3OD) : 27.3 (CH_3) , 58.3 $(C_{4,9})$, 73.4 $(C_{5,8})$, 82.5 $(C_{6,7})$, 110.5 (CMe₂), 161.2 (C₂); MS (NH₃): 246 (MH⁺), 263

(5*S*,6*R*,7*R*,8*S*)-5,6,7,8-Tetrahydroxy-6,7-*O*-methylethylidene-2-imino-1,3-diazonane (14). As described above for 13, the protected guanidino-sugar 14 was obtained as an hygroscopic solid on one hand, from the bis-epoxide 12 (92 μ mol scale, 97%), and on the other hand by *N*,*O*-debenzylation of 52 (97 μ mol scale, 80%). [α] +27 (α)

 $(M + NH_4^+)$.

^bBacillus stearothermophilus, pH 6.8.

cAlmond, pH 5.0.

dJack beans, pH 4.5.

^eBovine kidney, pH 5.5.

fInhibition constants (K_i in μ M) determined by the Lineweaver–Burk method.

0.9, MeOH); ¹H NMR (D₂O): 1.49 (s, 6H, CH₃), 3.42 (dd, 2H, $J_{4,5} = 5.8$, $J_{4,4'} = 15.6$ Hz, $H_{4,9}$), 3.64 (m, 2H, $H_{4',9'}$), 3.75–4.10 (m, 4H, $H_{5,6,7,8}$); ¹³C NMR (CD₃OD): 27.3 (CH₃), 55.5 (C_{4,9}), 72.4 (C_{5,8}), 82.3 (C_{6,7}), 110.6 (CMe₂), 161.7 (C₂); MS (NH₃): 246 (MH⁺).

1,6-Di-O-benzyl-3,4-O-methylethylidene-D-mannitol (15). In a flask equipped with a Dean-Stark, a suspension of 3,4-O-methylethylidene-D-mannitol (500 mg, 2.25 mmol) and dibutyl-tin oxide (1.15 g, 4.7 mmol, 2.1 equiv) in toluene (23 mL) was heated under reflux during 15 h. After concentration in vacuo, to a solution of the residue in toluene (11 mL), benzyl bromide (1.1 mL, 9 mmol, 4 equiv) and tetrabutylammonium iodide (805 mg, 2.25 mmol, 1 equiv) were added and the mixture was stirred at 70 °C for 15 h. Concentration in vacuo and subsequent chromatography of the resulting residue (EtOAc:cyclohexane, 3:7) afforded the diol 15 (777 mg, 86%) as an oil. $[\alpha] + 21$ (c 1.2, CHCl₃); ¹H NMR: 1.36 (s, 6H, CH₃), 3.57 (dd, 2H, $J_{1,2}$ =6.2, $J_{1,1'} = 9.7 \,\text{Hz}, \, H_{1,6}$), 3.71–3.85 (m, 4H, $H_{1',2,5,6'}$), 3.85– 3.95 (m, 2H, H_{3,4}), 4.57 (s, 4H, OCH₂Ph), 7,32 (s, 10H, CH_{Ph}); ¹³C NMR: 26.9 (CH₃), 71.7 (C_{1,6}), 72.0 (C_{2,5}), 73.5 (OCH₂Ph), 79.9 (C_{3.4}), 109.4 (CMe₂), 127.8, 128.4 (CH_{Ph}), 138.1 (C_{Ph}). Anal. calcd for C₂₃H₃₀O₆: C, 68.62; H, 7.52. Found: C, 68.77; H, 7.41.

(4R,5R,1'R,1"R)-4,5-Di-(2'-benzyloxy-1'-hydroxyethyl)-1,3,6-trioxepan (18) and 1,6-di-O-benzyl-3,4-O-methylene-D-mannitol (25). The diols 18 and 25 were obtained from the mixture of the tetrols 23 and 24 on a 0.22 mmol scale, as described above for the diol 15. Flash chromatography of the residue (cyclohexane:EtOAc, 6:4) respectively afforded 18 (40 mg, 30%) and 25 (10 mg, 8%).

18. $[\alpha]$ –2 (c 0.7, CHCl₃); ¹H NMR: 3.60–3.67 (m, 6H, H_{1,3,4,6}), 4.01 (m, 2H, H_{2,5}), 4.50, 4.55 (AB, 4H, J_{AB} = 11.0 Hz, OCH₂Ph), 4.78, 4.95 (2d, 4H, J = 6.0 Hz, OCH₂O), 4.95 (d, 2H, $J_{2,2}$ = 6.0 Hz, OCH₂O), 7.30 (s, 10H, CH_{Ph}); ¹³C NMR: 69.4 (C_{2,5}), 70.7 (C_{1,6}), 73.4 (OCH₂Ph), 83.0 (C_{3,4}), 91.9 (OCH₂O), 127.8, 128.5 (CH_{Ph}), 137.9 (C_{Ph}); MS (NH₃) 422 (M+NH₄⁺).

25. [α] +11 (c 0.4, CHCl₃); ¹H NMR: 3.57 (dd, 2H, $J_{1,2}$ =6.0, $J_{1,1'}$ =10.0 Hz, $H_{1,6}$), 3.71 (dd, 2H, $J_{1',2}$ =3.2, $J_{1,1'}$ =10.0 Hz, $H_{1',6'}$), 3.80 (m, 2H, $H_{2,5}$), 3.97 (m, 2H, $H_{3,4}$), 4.56 (s, 4H, OCH₂Ph), 4.93 (s, 2H, OCH₂O), 7.32 (s, 10H, CH_{Ph}); MS (NH₃): 392 (M+NH₄⁺).

(4S,5S,4'R,4"R)-4,5-Di-(2'-oxo-1',3'-dioxol-4'-yl)-1,3,6-trioxepan (21) and 1,2:5,6-di-O-carbonyl-3,4-O-methyl-ene-D-mannitol (22). Boron trifluoride etherate (215 μL) was dropwise added to a suspension of both the diol 20 (163 mg, 0.69 mmol) and paraformaldehyde (62 mg) in ethyl acetate (1.6 mL) at 0 °C. Then, the temperature was allowed to warm up to 20 °C and stirring was continued for 15 h. After addition of a saturated aqueous solution of NaHCO₃, the pH was adjusted to 8 by addition of solid NaHCO₃. Then the mixture was extracted with ethyl acetate and the combined organic layers were dried (MgSO₄), concentrated in vacuo and

chromatographied (acetone:dichloromethane, 5:95) to yield a mixture of the trioxepane **21** and of the methylene acetal **22** (75:25 according to 1 H NMR, 137 mg, \sim 71%) as a solid.

21. ¹H NMR: 3.78 (m, 2H, H_{4,5}), 4.43 (dd, 2H, $J_{4',5'}$ = 6.3, $J_{5',5''}$ = 9.0 Hz, H_{5'}), 4.55 (dd, 2H, $J_{5',5''}$ = 9.0, $J_{4',5''}$ = 8.2 Hz, H_{5''}), 4.78–4.86 (m, 2H, H_{4'}), 4.90 (d, 2H, $J_{2,2'}$ = 6 Hz, H_{2,7}), 5.07 (d, 2H, $J_{2,2'}$ = 6.0 Hz, H_{2,7}); MS (NH₃): 277 (MH⁺), 294 (M+NH₄⁺).

22. ¹H NMR: 4.11 (m, 2H, H_{3,4}), 4.42 (dd, 2H, $J_{1,2}$ = 5.3, $J_{1,1'}$ = 8.7 Hz, H_{1,6}), 4.58 (dd, 2H, $J_{1',2}$ = 9.5, $J_{1,1'}$ = 8.7 Hz, H_{1',6'}), 4.70–4.75 (m, 2H, H_{2,5}), 5.10 (s, 2H, OCH₂O); MS (NH₃): 264 (M+NH₄⁺).

(4R,5R,1'R,1"R) - 4,5 - Di - (1',2' - dihydroxyethyl) - 1,3,6-trioxepan (23) and 3,4-O-methylene-D-mannitol (24). A solution of the mixture of carbonates 21 and 22 (137 mg, \sim 0.5 mmol) in a 1:1 pyridine:water solution (26 mL) was heated under reflux during 1 h to yield a quantitative mixture of tetrols 23 and 24 (110 mg, \sim 75:25), after concentration in vacuo.

23. ¹H NMR (D₂O): 3.64–3.81 (m, 4H, H_{2',2"}), 3.86 (m, 2H, H_{1',1"}), 3.99 (m, 2H, H_{4,5}), 5.01 (d, 2H, J_{2,2'} = 8.0 Hz, H_{2',7}), 5.18 (d, 2H, J_{2,2'} = 8.0 Hz, H_{2',7'}).

24. ¹H NMR (D₂O): 3.64–3.81 (m, 4H, H_{1,6}), 3.86 (m, 2H, H_{2,5}), 4.18 (m, 2H, H_{3,4}), 5.08 (s, 2H, OCH₂O).

1,6-Di-*O***-benzyl-2,5-di-***O***-methane sulfonyl-3,4-***O***-methylethylidene-D-mannitol (26).** Methane sulfonyl chloride (150 μ L, 1.9 mmol, 3 equiv) was dropwise added to a solution of the diol **15** (250 mg, 0.62 mmol) and triethylamine (350 μ L, 2.5 mmol, 4 equiv) in dichloromethane (1 mL) at 0 °C. After stirring at 20 °C for 5 min, water was added and the mixture was extracted with dichloromethane. The combined organic layers were dried (MgSO₄) and concentrated in vacuo to quantitatively yield the dimesylate **26** (345 mg). ¹H NMR 1.38 (s, 6H, CH₃), 3.03 (s, 6H, SO₃Me), 3.70 (dd, 2H, $J_{1,2}$ = 6.9, $J_{1,1'}$ = 11.3 Hz, $H_{1,6}$), 3.85 (dd, 2H, $J_{1',2}$ = 3.2, $J_{1,1'}$ = 11.3 Hz, $H_{1',6'}$), 4.33 (m, 2H, $H_{3,4}$), 4.51, 4.57 (AB, 4H, J_{AB} = 11.7 Hz, OCH₂Ph), 4.86 (m, 2H, $H_{2,5}$), 7.30 (m, 10H, CH_{Ph}).

1,3,4,6-Tetra-*O***-benzyl-2,5-di-***O***-methane sulfonyl-D-mannitol (27).** The dimesylate **27** (130 mg) was quantitatively obtained from the diol **16** as described above for the dimesylate **26** on a 0.18 mmol scale. 1 H NMR 2.96 (s, 6H, SO₂CH₃), 3.80 (dd, 2H, $J_{1,2}$ = 7.0, $J_{1,1'}$ = 11.0 Hz, H_{1,6}), 3.92 (dd, 2H, $J_{1',2}$ = 3.0, $J_{1,1'}$ = 11.0 Hz, H_{1',6'}), 4.03 (m, 2H, H_{3,4}), 4.45, 4.52 (AB, 4H, J_{AB} = 12.0 Hz, OCH₂Ph), 4.62, 4.68 (AB, 4H, J_{AB} = 11.0 Hz, OCH₂Ph), 4.93 (m, 2H, H_{2.5}), 7.32 (m, 20H, CH_{Ph}).

1,3:4,6-Di-*O***-benzylidene-2,5-di-***O***-methanesulfonyl-D-mannitol (28).** The dimesylate **28** (157 mg, 79%) was obtained from the diol **17** as described above for the

dimesylate **26** on a 0.28 mmol scale. ¹H NMR: 3.03 (s, 6H, SO₂CH₃), 3.84 (dd, 2H, $J_{1,2}$ =10.5, $J_{1,1'}$ =10.6 Hz, H_{1,6}), 4.13 (d, 1H, $J_{2,3}$ =9.0 Hz, H_{3,4}), 4.55 (dd, 2H, $J_{1',2}$ =5.5, $J_{1',1}$ =10.6 Hz, H_{1',6'}), 5.05 (m, 2H, H_{2,5}), 5.50 (s, 2H, OCHPh), 7.35 (m, 6H, CH_{Ph}), 7.47 (m, 4H, CH_{Ph}).

1,3,4,6-Tetra-*O*-benzyl-2,5-anhydro-D-glucitol (29). Diisopropyl azodicarboxylate (55 µL, 0.27 mmol, 3 equiv) was dropwise added to a solution of both the diol 16 (50 mg, 0.09 mmol) and triphenylphosphine (55 mg, 0.27 mmol, 3 equiv) in toluene (1 mL) at 0 °C. After stirring at 0 °C for 1 h, the reaction mixture was concentrated in vacuo and the residue was chromatographied (EtOAc:cyclohexane, 1:9) to yield the tetrahydrofuran 29 (44 mg, 91%). [α] +21 (c 1.2, CHCl₃) {lit.⁶⁷ [α]_D²⁰ +24.9 $(c 4.28, CHCl_3)$; ¹H NMR: 3.54 (dd, 1H, $J_{1,2} = 6.8$, $J_{1,1'} = 9.9 \text{ Hz}, H_1$, 3.65 (dd, 1H, $J_{1',2} = 5.7, J_{1,1'} = 9.9 \text{ Hz},$ $H_{1'}$), 3.73 (dd, 1H, $J_{5,6} = 6.0$, $J_{6,6'} = 11.3$ Hz, H_6), 3.79 (dd, 1H, $J_{5,6'} = 5.3$, $J_{6,6'} = 11.3$ Hz, $H_{6'}$), 3.96 (br.d, 1H, $J_{2,3} = 3.0 \text{ Hz}, H_3$, 3.98 (br.d, 1H, $J_{4,5} = 3.9 \text{ Hz}, H_4$), 4.12 (ddd, 1H, $J_{2,3} = 3.0$, $J_{1',2} = 5.7$, $J_{1,2} = 6.8$ Hz, H₂), 4,26 (ddd, 1H, $J_{4,5} = 3.9$, $J_{5,6}' = 5.3$, $J_{5,6} = 6.0$ Hz, H₅), 4.39, 4.62 (AB, 2H, $J_{AB} = 12.0$ Hz, OCH₂Ph), 4.61-4.47 (m, 6H, OCH₂Ph), 7.29 (br.s, 20H, CH_{Ph}); ¹³C NMR: 68.3, 70.5 (C_{1,6}), 71.5, 73.4 (OCH₂Ph), 80.1, 82.8, 83.8 (C_{2,3,4,5}), 127.6, 128.4, 129.7, 133.5 (CH_{Ph}), 137.9 (C_{Ph}); MS (NH₃): $542 (M + NH_4^+)$.

(1*R*,7*R*,8*S*,10*R*)-8,10-Dibenzyloxy methyl-2,4,6,9-tetra-oxa-bicyclo[5.3.0] decane (30). The tetrahydrofuran 30 was obtained from the diol 18 as described above for the tetrahydrofuran 29 on a 0.1 mmol scale. After flash chromatography (cyclohexane:EtOAc, 8:2), the compound 30 (26 mg, 68%) was obtained as an oil. [α] –4 (*c* 1.0, CHCl₃); ¹H NMR: 3.53 (dd, 1H, $J_{10,\text{CHOBn}}$ =4.8, $J_{\text{CH}_2\text{OBn}}$ =10.6 Hz, CH₂OBn), 3.60–3.69 (m, 3H, CH₂OBn), 4.03 (m, 1H, H₁₀), 4.25–4.36 (m, 2H, H_{1,8}), 4.45 (dd, 1H, $J_{7,8}$ =8.3, $J_{1,7}$ =8.1 Hz, H₇), 4.53 (s, 2H, OCH₂Ph), 4.58 (s, 2H, OCH₂Ph), 4.82 (d, 1H, $J_{3,3'}$ =6.2 Hz, H₃), 4.95 (d, 1H, $J_{5,5'}$ =6.2 Hz, H₅), 5.11 (d, 2H, $J_{3,3'}$ = $J_{5,5'}$ =6.2 Hz, H_{3,5}), 7.31 (s, 10H, CH_{Ph}); ¹³C NMR: 69.7, 69.9 (CH₂OBn), 73.5 (OCH₂Ph), 76.4, 77.5, 80.1, 82.3 (C_{1,7,8,10}), 95.5, 95.9 (C_{3,5}), 127.5, 127.6, 129.7, 133.0 (CH_{Ph}), 138.3 (C_{Ph}); MS (NH₃): 404 (M+NH₄+).

2,5-Diazido-1,6-di-O-benzyl-2,5-dideoxy-3,4-O-methylethylidene-L-iditol (31). A solution of the dimesylate 26 (345 mg, 0.62 mmol) and sodium azide (405 mg, 6.2 mmol, 10 equiv) in DMF (7.5 mL) was heated at 120 °C during 15 h. After concentration in vacuo and addition of water, the mixture was extracted with dichloromethane. The combined organic layers were dried (MgSO₄), concentrated in vacuo and chromatographied (cyclohexane:dichloromethane, 2:8) to yield the diazido compound 31 (160 mg, 57%) as a colorless oil. [α] +67 (c 1.7, CHCl₃); ¹H NMR: 1.41 (s, 6H, CH₃), 3.67 (dd, 2H, $J_{1,2} = 4.7$, $J_{1,1'} = 10$ Hz, $H_{1,6}$), 3.75 (dd, 2H, $J_{1'2} = 7.8$, $J_{1,1'} = 10$ Hz, $H_{1',6'}$), 4.13 (br.s, 2H, H_{3.4}), 4.46 (m, 2H, H_{2.5}), 4.55 (s, 4H, OCH₂Ph), 7.34 (s, 10H, CH_{Ph}); ¹³C NMR: 26.9 (CH₃), 60.1 (C_{2.5}), 69.9, 73.6 (C_{1,6}, OCH₂Ph), 77.1 (C_{3,4}), 110.5 (CMe₂), 127.8, 127.9, 128.5 (CH_{Ph}), 137.5 (C_{Ph}); MS (NH₃): 425 (MH^+-N_2) , 470 $(M^++NH_4^+)$. Anal. calcd for $C_{23}H_{28}O_4N_6$: C, 61.03; H, 6.24; N, 18.58. Found: C, 61.14; H, 6.18; N, 18.56.

2,5-Diamino-1,6-di-O-benzyl-2,5-dideoxy-3,4-O-methylethylidene-L-iditol (32). Palladium (10%) on charcoal (125 mg) in ethanol (3 mL) was completely hydrogenated under 1 atm of H₂ at 20 °C prior to the addition of the diazido derivative 31 (189 mg, 0.42 mmol) in ethanol (2 mL). After 2 h, the catalyst was removed through a celite pad and the filtrate was concentrated in vacuo and purified by flash chromatography (CH₂Cl₂:MeO-H:ammonium hydroxide, 94:3:3) to yield the diamine 32 $(146 \,\mathrm{mg}, \,87\%)$ as an oil. [α] +14 (c 0.8, CHCl₃); ¹H NMR: 1.38 (s, 6H, CH₃), 2.93 (m, 2H, H_{2,5}), 3.80 (dd, 2H, $J_{1,2} = 7.3$, $J_{1,1'} = 9.2$ Hz, $H_{1,6}$), 3.49 (dd, 2H, $J_{1',2} = 5$, $J_{1,1'} = 9.2 \,\text{Hz}, \, H_{1',6'}$, 4.06 (br.s, 2H, $H_{3,4}$), 4.49 (s, 4H, OCH₂Ph), 7.30 (s, 10H, CH_{Ph}); ¹³C NMR (CDCl₃): 19.7 (CH₃), 51.9 (C_{2.5}), 73.3, 73.8 (C_{1.6}, OCH₂Ph), 78.2 $(C_{3.4})$, 108.8 (CMe₂), 127.7, 128.4 (CH_{Ph}), 138.1 (C_{Ph}).

(4*S*,5*R*,6*R*,7*S*)-4,7-Dibenzyloxymethyl-5,6-dihydroxy-5,6-O-methylethylidene-1,3-diazepan-2-thione (33). Carbon disulfide (118 µL, 1.96 mmol, 2 equiv) was added to a solution of the diamine 32 (391 mg, 0.98 mmol) in pyridine (800 µL). After stirring at 60 °C for 15 h, the pH was adjusted to 2–3 by the addition of a 1 M aq HCl solution. The mixture was extracted with dichloromethane and the combined organic layers were washed with a 1 M aq NaOH solution, dried (MgSO₄) and concentrated in vacuo to yield the thiourea 33 (372 mg, 86%) as a yellow foam. $[\alpha] + 21$ (c 1.0, CHCl₃); ¹H NMR: 1.35 (s, 6H, CH₃), 3.30–3.80 (m, 4H, CH₂OBn), 3.80-4.20 (m, 4H, $H_{4,5,6,7}$), 4.20-4.60 (m, 4H, OCH₂Ph), 7.24 (m, 10H, CH_{Ph}); 13 C NMR: 27.0 (CH₃), 53.0 (C_{4,7}), 69.5 (C_{5.6}), 73.1 (CH₂OBn, OCH₂Ph), 110.2 (CMe₂), 127.8, 128.4 (CH_{Ph}), 136.3 (C_{Ph}), 184.0 (C₂); IR (neat): 1230 ($\nu_{C=S}$); HRMS (NH₃): for $C_{24}H_{31}N_2O_4S$ (MH⁺) calcd 443.2004, found 443.1983.

(4*R*,5*R*,6*R*,7*R*)-4,7-Dibenzyloxymethyl-5,6-dihydroxy-2-methylthio-4,5,6,7-tetrahydro-1,3-diazepinium *para*-toluene sulfonate (34). A solution of the thiourea 33 (40 mg, 0.09 mmol) and methyl *para*-toluenesulfonate (24 μL, 0.16 mmol, 1.7 equiv) in methanol (200 μL) was heated under reflux during 30 min. After concentration in vacuo, the resulting oil was triturated with diethylether to yield the isothiourea 34 (47 mg, 88%) as an hygroscopic solid. [α] +109 (c 0.4, CHCl₃); ¹H NMR: 2.29 (br.s, 3H, SCH₃), 2.44 (br.s, 3H, ArCH₃), 3.80 (m, 6H, H_{4,7}, CH₂OBn), 4.40 (m, 6H, H_{5,6}, OCH₂Ph), 7.09 (br.s, 2H, CH_{Ar}); ¹³C NMR: 15.4 (SCH₃), 21.3 (ArCH₃), 60.6 (C_{4,7}), 69.6 (CH₂OBn), 71.6 (C_{5,6}), 73.4 (OCH₂Ph), 126.0, 127.8, 128.0, 128.5, 128.9, 129.9 (CH_{Ar}), 137.5, 140.3, 141.9 (C_{Ar}); MS (FAB+): 417 (MH⁺).

(4S,5R,6R,7S)-4,7-Dibenzyloxymethyl-5,6-dihydroxy-5,6 -*O*-methylethylidene-2-(*N*-benzyl)-imino-1,3-diazepane (35). Benzylamine (57 μ L, 0.52 mmol, 1.1 equiv) and triethylamine (63 μ L, 0.94 mmol, 2 equiv) were successively added to a solution of both the thiourea 33 (208 mg, 0.47 mmol) and mercuric chloride (160 mg,

0.61 mmol, 1.3 equiv) in dichloromethane (6.5 mL). After stirring at 20 °C during 15 h, mercuric salts were removed by filtration through a celite pad and washed with dichloromethane. The combined organic layers were successively washed with a 1 M aq HCl solution and a 1 M aq NaOH solution then dried (MgSO₄) and concentrated in vacuo to yield the guanidine **35** (220 mg, 91%). [α] +32 (c 0.5, CHCl₃); 1 H NMR: 1.25 (br.s, 6H, CH₃), 3.30–3.70 (m, 4H, CH₂OBn), 3.70–4.00 (m, 6H, H_{4,5,6,7}, NCH₂Ph), 4.00–4.38 (m, 4H, OCH₂Ph), 7,24 (br.s, 15H, CH_{Ph}); 13 C NMR: 27.1 (CH₃), 53.4 (C_{4,7}), 56.3 (C5,6), 71.5, 73.0 (NCH₂Ph, CH₂OBn, OCH₂Ph), 109.5 (CMe₂), 127.7, 128.4 (CH_{Ph}), 138.0 (C_{Ph}); IR (neat): 1625 (ν _{C=N}); HRMS (NH₃) for C₃₁H₃₈N₃O₄: (MH⁺) calcd 516.2862, found 516.2845.

(4S,5R,6R,7S)-5,6-Dihydroxy-4,7-dihydroxy methyl-5,6-O-methylethylidene-2-imino-1,3-diazepane (36). A solution of the benzylated guanidine 35 (191 mg, 0.42 mmol) in a minimum of anhydrous THF was poured into liquid ammonia (10 mL). Then, sodium was added until persistance of a dark blue color and the mixture was stirred in refluxing ammonia during 6h. After careful addition of solid ammonium chloride until decoloration, ammonia was evaporated and the solid residue was washed with dichloromethane and extracted with absolute ethanol. The combined alcoholic layers were concentrated in vacuo. Neutralization of the resulting residue by ethanolic elution through an Amberlite® IRA 400 (OH-) packed column followed by concentration in vacuo afforded the guanidine 36 (67 mg, 74%) as an hygroscopic solid. ¹H NMR (D₂O): 1.60 (m, 6H, CH₃), 3.70-4.30 (m, 8H, $H_{4,5,6,7}$, CH_2OH); ^{13}C NMR (D_2O): 28.9 (CH₃), 57.0 (C_{4.7}), 64.6 (CH₂OH), 79.3 (C_{5.6}), 113.4 (CMe_2) , 160.2 (C_2) .

1,6-Di-O-benzyl-3,4-O-methylethylidene-L-iditol (37). Benzylic alcohol (4.2 mL, 40.3 mmol, 2.5 equiv) was added to a suspension of sodium hydride (851 mg, 35.5 mmol, 2.2 equiv) in DMF (32 mL) and the reaction mixture was stirred at 20 °C during 3 h. Then, a solution of the bis-epoxide 12³⁹ (3 g, 16.11 mmol) in a minimum of DMF was dropwise added at 0 °C. After stirring at 20 °C during 20 h, methanol was added and the reaction mixture was concentrated in vacuo prior to the addition of water. The mixture was then extracted with dichloromethane and the combined organic layers were dried (MgSO₄), concentrated in vacuo and chromatographied (cyclohexane:AcOEt, 70:30) to yield the diol 37 (3.7 g, 57%) as an oil. [α] -19 (c 5.1, CH₂Cl₂); ¹H NMR: 1.43 (s, 6H, CH₃), 3.51 (dd, 2H, $J_{1,2} = 5.5$, $J_{1,1'} = 10.0$ Hz, $H_{1.6}$), 3.56 (dd, 2H, $J_{1',2} = 6.0$, $J_{1,1'} = 10.0$ Hz, $H_{1',6'}$), 3.78 (m, 2H, H_{2.5}), 4.11 (m, 2H, H_{3.4}), 4.52 (s, 4H, OCH₂Ph), 7.33 (s, 10H, CH_{Ph}). Anal. calcd for $C_{23}H_{30}O_6$: C, 68.64; H, 7.51. Found: C, 68.54; H, 7.50.

1,6-Di-*O***-benzyl-2,5-di-***O***-methane sulfonyl-3,4-***O***-methylethylidene-**L**-iditol (38).** The dimesylate **38** (2.57 g) was quantitatively obtained from the diol **37** as described above for the dimesylate **26** on a 4.59 mmol scale. 1 H NMR: 1.40 (s, 6H, CH₃), 3.02 (s, 6H, SO₃CH₃), 3.70 (dd, 2H, $J_{1,2}$ = 3.3, $J_{1,1'}$ = 11.0 Hz, $H_{1,6}$), 3.87 (dd, 2H, $J_{1,2}$ = 8.5, $J_{1,1'}$ = 11.0 Hz, $H_{1',6'}$), 4.13 (m, 2H, $H_{3,4}$), 4.48,

4.54 (AB, 4H, $J_{AB} = 11.5$ Hz, OCH₂Ph), 4.92 (m, 2H, H_{2.5}), 7.33 (m, 10H, CH_{Ph}).

2,5-Diazido-1,6-di-*O***-benzyl-2,5-dideoxy-3,4-***O***-methylethylidene-D-mannitol** (39). The diazido compound 39 was obtained from the dimesylate 38 as described above for the diazide 31 on a 4.6 mmol scale. After flash chromatography (dichloromethane:cyclohexane, 8:2), the product 39 (1.3 g, 63% yield) was obtained as an oil. [α] +8 (c 0.8, CHCl₃); 1 H NMR: 1.35 (s, 6H, CH₃), 3.61 (dd, 2H, $J_{1,1'}$ =8.3, $J_{1,2}$ =9.4 Hz, $H_{1,6}$), 3.69–3.75 (m, 2H, $H_{2,5}$), 3.81 (dd, 2H, $J_{1',2}$ =3.0, $J_{1,1'}$ =8.3 Hz, $H_{1',6'}$), 4.01 (m, 2H, $H_{3,4}$), 4.57 (s, 4H, OCH₂Ph), 7.33 (s, 10H, CH_{Ph}); 13 C NMR: 27.4 (CH₃), 63.1 (C_{2,5}), 70.0, 73.6 (C_{1,6}, OCH₂Ph), 78.0 (C_{3,4}), 110.7 (CMe₂), 126.5, 127.8, 128.5 (CH_{Ph}), 137.6 (C_{Ph}); IR (neat): 2100 (v_{N₃}). Anal. calcd for C₂₃H₂₈O₄N₆: C, 61.05; H, 6.24; N, 18.57. Found: C, 61.02; H, 6.35; N, 18.47.

2,5-Diamino-1,6-di-*O***-benzyl-2,5-dideoxy-3,4-***O***-methylethylidene-D-mannitol** (**40**). The diamine **40** was obtained from the diazido compound **39** as described above for the diamine **32** on a 0.89 mmol scale. After flash chromatography (CH₂Cl₂:MeOH:ammonium hydroxide, 94:3:3), the compound **40** (280 mg, 79%) was obtained as an oil. [α] + 7 (c 0.7, CHCl₃); ¹H NMR: 1.33 (s, 6H, CH₃), 3.07 (m, 2H, H_{2,5}), 3.46 (dd, 2H, $J_{1,2}$ = 7.1, $J_{1,1'}$ = 9.1 Hz, H_{1,6}), 3.69 (dd, 2H, $J_{1',2}$ = 3.5, $J_{1,1'}$ = 9.1 Hz, H_{1',6}), 3.85 (m, 2H, H_{3,4}), 4.52 (s, 4H, OCH₂Ph), 7.32 (s, 10H, CH_{Ph}); ¹³C NMR: 27.3 (CH₃), 54.0 (C_{2,5}), 72.6, 73.3 (C_{1,6}, OCH₂Ph), 81.1 (C_{3,4}), 108.9 (CMe₂), 127.7, 128.4 (CH_{Ph}), 138.3 (C_{Ph}).

1,6-Diazido-1,6-dideoxy-3,4-O-methyl ethylidene-L-iditol (41). A solution of the bis-epoxide 12^{39} (688 mg, 3.7 mmol), ammonium chloride (870 mg, 16.3 mmol, 4.4 equiv) and sodium azide (2.4 g, 36.9 mmol, 10 equiv) in methanol:water 8:1 (15 mL) was stirred at 20 °C during 15 h. After concentration in vacuo, water was added and the mixture was extracted with dichloromethane. The combined organic layers were dried (MgSO₄), concentrated in vacuo and chromatographied (dichloromethane:ether, 80:20) to yield the diazido-diol 41 (835 mg, 83%) as an oil. $[\alpha]$ -4 (c 1.0, CH₂Cl₂); ¹H NMR: 1.43 (s, 6H, CH₃), 3.35 (dd, 2H, $J_{1.2} = 5.2$, $J_{1.1'} =$ 12.6 Hz, $H_{1,6}$), 3.45 (dd, 2H, $J_{1',2} = 6.9$, $J_{1,1'} = 12.6$ Hz, $H_{1',6'}$), 3.73 (m, 2H, $H_{2,5}$), 4.04 (s, 2H, $H_{3,4}$); ¹³C NMR: 27.0 (CH₃), 54.2 (C_{1,6}), 68.9 (C_{2,5}), 77.0 (C_{3,4}), 110.3 (CMe₂); IR (neat): 2100 (v_{N_2}). Anal. calcd for C₉H₁₆N₆O₄: C, 39.70; H, 5.92; N, 30.87. Found: C, 39.83; H, 3.17; N, 30.11.

1,6-Diazido-2,5-di-O-benzyl-1,6-dideoxy-3,4-O-methylethylidene-L-iditol (42). A solution of the diol 41 (748 mg, 2.75 mmol) in THF (4.5 mL) was dropwise added to a suspension of sodium hydride (195 mg, 8.1 mmol, 3 equiv) in THF (4.5 mL) at 0 °C. After stirring at 20 °C for 3 h, benzyl bromide (4 mL, 8.1 mmol, 3 equiv) was added and the mixture was stirred at 20 °C for an additional 15 h. Then, methanol was added and the reaction mixture was concentrated in vacuo. After addition of water and extraction with dichloromethane, the combined organic layers were washed with brine,

dried (MgSO₄), and concentrated in vacuo. Flash chromatograpy (dichloromethane:cyclohexane, 8:2) afforded the compound **42** (957 mg, 77%) as an oil. [α] + 34 (c1.0, CHCl₃); ¹H NMR: 1.37 (s, 6H, CH₃), 3.41 (br.s, 6H, H_{1,2,5,6}), 4.04 (s, 2H, H_{3,4}), 4.51, 4. 71 (AB, 4H, J_{AB} = 11.7 Hz, OCH₂Ph), 7.32 (br.s, 10H, CH_{Ph}); ¹³C NMR: 26.9 (CH₃), 51.8 (C_{1,6}), 73.4 (OCH₂Ph), 76.0, 76.2 (C_{2,3,4,5}), 109.5 (CMe₂), 128.1, 128.2, 128.5 (CH_{Ph}), 137.5 (C_{Ph}); IR (neat) 2100 (v_{N_3}). Anal. calcd for C₂₃H₂₈N₃O₄: C,61.0; H, 6.24; N, 18.57. Found: C, 61.00; H, 6.25; N, 18.43.

1,6-Diamino-2,5-di-*O***-benzyl-1,6-dideoxy-3,4-***O***-methylethylidene-L-iditol (43).** The diamine **43** was obtained from the diazido compound **42** as described above for the diamine **32** on a 0.85 mmol scale. After flash chromatography (CH₂Cl₂:MeOH:ammonium hydroxide, 90:7:3), the compound **43** (240 mg, 70%) was obtained as an oil. [α] +12 (c 0.4, CHCl₃); 1 H NMR: 1.40 (s, 6H, CH₃), 2.77 (dd, 2H, $J_{1,2}$ =5.9, $J_{1,1'}$ =13.3 Hz, H_{1,6}), 2.89 (dd, 2H, $J_{1',2}$ =5.0, $J_{1,1'}$ =13.3 Hz, H_{1',6'}), 3.34 (m, 2H, H_{2,5}), 4.14 (br.s, 2H, H_{3,4}), 4.57, 4.64 (AB, 4H, J_{AB} =11.8 Hz, OCH₂Ph), 7.30 (br.s, 10H, CH_{Ph}); 13 C NMR: 27.1 (CH₃), 42.3 (C_{1,6}), 72.7 (OCH₂Ph), 77.7, 78.8 (C_{2,3,4,5}), 109.1 (CMe₂), 127.8, 128.0, 128.4 (CH_{Ph}), 138.2 (C_{Ph}).

1,6-Diazido-1,6-dideoxy-3,4-*O*-methyl ethylidene-D-mannitol (45). A solution of the ditosylate 44^{39} (5 g, 9.5 mmol) and sodium azide (2.5 g, 38 mmol, 4 equiv) in DMF (38 mL) was heated at 70 °C during 3 h. After concentration in vacuo and addition of water, the mixture was extracted with dichloromethane. The combined organic layers were dried (MgSO₄), concentrated in vacuo and chromatographied (dichloromethane:ether, 90:10) to yield the diazido-diol **45** (2 g, 75%) as an oil. [α] +45 (c 2.0, CH₂Cl₂); 1 H NMR: 1.35 (s, 6H, CH₃), 3.44 (dd, 2H, $J_{1,2}$ =4.6, $J_{1,1'}$ =12.2 Hz, $H_{1,6}$), 3.64 (d, 2H, $J_{1,1'}$ =12.2 Hz, $H_{1,6'}$), 3.78 (m, 4H, $H_{2,3,4,5}$); 13 C NMR: 26.7 (CH₃), 54.2 (C_{1,6}), 72.2 (C_{2,5}), 80.1 (C_{3,4}), 109.9 (CMe₂); IR (neat) 2100 (v_{N3}). Anal. calcd for C₉H₁₆N₆O₄: C, 39.70; H, 5.92; N, 30.87. Found: C, 39.72; H, 6.23; N, 29.93.

1,6-Diazido-2,5-di-*O***-benzyl-1,6-dideoxy-3,4-***O***-methylethylidene** - **D** - **mannitol** (**46**). The dibenzyl-ether **46** was obtained from the diol **45** as described above for the dibenzyl-ether **42** on a 1.78 mmol scale. After flash chromatography (dichloromethane:cyclohexane, 8:2), the compound **46** (223 mg, 81%) was obtained as an oil. 1 H NMR: 1.34 (s, 6H, CH₃), 3.39 (dd, 2H, $J_{1,2}$ = 5.8, $J_{1,1'}$ = 13.2 Hz, $H_{1,6}$), 3.49 (dd, 2H, $J_{1',2}$ = 3.3, $J_{1,1'}$ = 13.2 Hz, $H_{1',6'}$), 3.61 (m, 2H, $H_{2,5}$), 4.06 (m, 2H, $H_{3,4}$), 4.53, 4.66 (AB, 4H, J_{AB} = 11.5 Hz, OCH₂Ph), 7.29 (m, 10H, CH_{Ph}); 13 C NMR: 27.2 (CH₃), 51.1 (C_{1,6}), 72.8 (OCH₂Ph), 78.3, 79.3 (C_{2,3,4,5}), 110.2 (CMe₂), 128.0, 128.5 (CH_{Ph}), 137.5 (C_{Ph}). Anal. calcd for C₂₃ H_{28} N₆O₄: C, 61.05; H, 6.24; N, 18.57. Found: C, 61.21; H, 6.16; N, 18.51.

1,6-Diamino-2,5-di-*O***-benzyl-1,6-dideoxy-3,4-***O***-methylethylidene-D-mannitol (47).** The diamine **47** was obtained from the diazido compound **46** as described above for the diamine **32** on a 0.29 mmol scale. After flash chromatography (CH₂Cl₂:MeOH:ammonium hydroxide, 90:7:3),

the compound **47** (94 mg, 82%) was obtained as an oil. [α] + 16 (c 1.0, CHCl₃); 1 H NMR: 1.36 (s, 6H, CH₃), 2.93 (m, 4H, H_{1.6}), 3.51 (m, 2H, H_{2.5}), 4.14 (m, 2H, H_{3.4}), 4.56 (s, 4H, OCH₂Ph), 7.27 (br.s, 10H, CH_{Ph}); 13 C NMR: 27.3 (CH₃), 41.2 (C_{1.6}); 72.1 (OCH₂Ph), 78.7, 81.4 (C_{2,3,4,5}), 109.7 (CMe₂), 127.9, 128.4 (CH_{Ph}), 138.2 (C_{Ph}).

(4*R*,5*R*,6*R*,7*R*)-4,7-Dibenzyloxymethyl-5,6-dihydroxy-5,6-*O*-methylethylidene-1,3-diazepan-2-thione (48). The thiourea 48 (125 mg, 81%) was obtained as a yellow foam from the diamine 40 on a 0.35 mmol scale, according to the procedure described above for the thiourea 33. [α] +40 (c 0.4, CHCl₃); 1 H NMR: 1.34 (s, 6H, CH₃), 3.44 (m, 6H, H_{4.7}, CH₂OBn), 3.74 (m, 2H, H_{5.6}), 4.55 (s, 4H, OCH₂Ph), 7.33 (s, 10H, CH_{Ph}); 13 C NMR: 26.8 (CH₃), 56.7 (C_{4.7}), 69.0, 73.4 (CH₂OBn, OCH₂Ph), 78.9 (C_{5.6}), 111.6 (CMe₂), 127.0, 128.6 (CH_{Ph}), 137.2 (C_{Ph}), 187.3 (C₂); IR (neat) 1230 (v_{C=S}); HRMS (NH₃) for C₂₄H₃₁ N₂O₄S: (MH⁺) calcd 443.2004, found 443.2003.

(4*R*,5*R*,6*R*,7*R*)-4,7-Dibenzyloxymethyl-5,6-dihydroxy-5,6-*O*-methylethylidene-2-(*N*-benzyl)-imino-1,3-diazepane (49). The guanidine 49 (45 mg, 78%) was obtained as a yellow foam from the thiourea 48 on a 0.11 mmol scale, according to the procedure described above for the guanidine 35. [α] + 37 (c 0.4, CHCl₃); ¹H NMR: 1.33 (s, 6H, CH₃), 3.42–3.72 (m, 6H, H_{4,7}, CH₂OBn), 3.80 (m, 2H, H_{5,6}), 4.18–4.41 (m, 2H, NCH₂Ph), 4.54 (m, 4H, OCH₂Ph), 7.14–7.25 (m, 15H, CH_{Ph}); ¹³C NMR: 26.7 (CH₃), 55.5 (C_{4,7}), 68.4, 73.5 (NCH₂Ph, CH₂OBn, OCH₂Ph), 78.2 (C_{5,6}), 111.6 (CMe₂), 127.0, 128.0, 128.3, 128.5, 129.0, 129.1 (CH_{Ph}), 135.2, 137.4 (C_{Ph}), 159.8 (C₂); IR (neat): 1635 (ν_{CN}); HRMS (NH₃) for C₃₁H₃₈N₃O₄: (MH⁺) calcd 516.2862, found 516.2872.

(4*R*,5*R*,6*R*,7*R*)-5,6-Dihydroxy-4,7-dihydroxy methyl-5,6-*O*-methylethylidene-2-imino-1,3-diazepane (50). The guanidino-sugar 50 (21 mg, 75%) was obtained as an hygroscopic solid from the guanidine 49 on a 0.13 mmol scale, according to the procedure described above for the guanidino-sugar 36. 1 H NMR (D₂O): 1.52 (s, 6H, CH₃), 3.55 (m, 2H, H_{4,7}), 3.79 (dd, 2H, $J_{4,\text{CHOH}}$ = 5.6, $J_{\text{CH}_{2}\text{OH}}$ = 12.1 Hz, CH₂OH), 3.91–3.99 (m, 4H, H_{5,6}, CH'OH); 13 C NMR (D₂O): 28.7 (CH₃), 59.2 (C_{4,7}), 63.4 (CH₂OH), 80.4 (C_{5,6}), 115.4 (CMe₂), 162.8 (C₂).

(5*S*,6*R*,7*R*,8*S*)-5,8-Dibenzyloxy-6,7-dihydroxy-6,7-*O*-methylethylidene-1,3-diazonan-2-thione (51). The thiourea 51 (335 mg) was quantitatively obtained as a yellow foam from the diamine 43 on a 0.77 mmol scale according to the procedure described above for thiourea 33. [α] +21 (*c* 1.1, CHCl₃); ¹H NMR: 1.36 (br.s, 6H, CH₃), 3.43 (m, 4H, H_{4,9}), 3.65 (m, 2H, H_{5,8}), 4.01 (br.s, 2H, H_{6,7}), 4.43, 4.58 (AB, 4H, J_{AB} = 11.8 Hz, OCH₂Ph), 7.25 (s, 10H, CH_{Ph}); ¹³C NMR: 27.0 (CH₃), 44.9 (C_{4,9}), 72.3 (OCH₂Ph), 73.8, 77.4 (C_{5,6,7,8}), 109.5 (CMe₂), 128.3 (CH_{Ph}), 137.5 (C_{Ph}), 182.6 (C₂); IR (neat): 1215 (ν_{C=S}); HRMS (NH₃) for C₂₄H₃₁N₂O₄S: (MH⁺) calcd 443.2004, found 443.2003.

(5*S*,6*R*,7*R*,8*S*)-5,8-Dibenzyloxy-6,7-dihydroxy-6,7-*O*-methylethylidene - 2 - (*N*-benzyl) - imino - 1,3 - diazonane (52). The guanidine 52 (55 mg, 94%) was obtained as a yellow

foam from the thiourea **51** on a 0.11 mmol scale according to the procedure described above for the guanidine **35**. [α] +21 c 1.1, CHCl₃); ¹H NMR: 1.25 (br.s, 6H, CH₃), 3.15–3.82 (m, 8H, H_{4,5,8,9}, NCH₂Ph), 3.8–4.30 (m, 4H, H_{6,7}), 4.30–4.80 (m, 4H, OCH₂Ph), 7.20 (br.s, 15H, CH_{Ph}); ¹³C NMR: 27.0 (CH₃), 43.6 (C_{4,9}, NCH₂Ph), 73.5 (OCH₂Ph), 77.7 (C_{5,6,7,8}), 109.9 (CMe₂), 128.1, 128.5 (CH_{Ph}), 137.6 (C_{Ph}), 156 (C₂); IR (neat): 1630 (ν _{C=N}); HRMS (NH₃) for C₃₁H₃₈N₃O₄: (MH⁺) calcd 516.2862, found 516.2872.

(5*R*,6*R*,7*R*,8*R*)-5,8-Dibenzyloxy-6,7-dihydroxy-6,7-*O*-methylethylidene-1,3-diazonan-2-thione (53). The thiourea 53 (145 mg) was quantitatively obtained as a yellow foam from the diamine 47 on a 335 μmol scale, according to the procedure described above for the thiourea 33. [α] +20 (c 0.4, CHCl₃); 1 H NMR: 1.34 (s, 6H, CH₃), 3.65 (m, 6H, H_{4,5,8,9}), 4.06 (m, 2H, H_{6,7}), 4.54 (m, 4H, OCH₂Ph), 7.26 (br.s, 10H, CH_{Ph}); 13 C NMR: 27.0 (CH₃), 44.3 (C_{4,9}), 72.3 (OCH₂Ph), 77.9, 78.4 (C_{5,6,7,8}), 110.1 (CMe₂), 127.9, 128.2, 129.4 (CH_{Ph}), 137.7 (C_{Ph}), 182.7 (C₂); IR (neat): 1210 (v_{C=S}); HRMS (NH₃) for C₂₄H₃₁N₂O₄S: (MH⁺) calcd 443.2004, found 443.2009.

(5*R*,6*R*,7*R*,8*R*)-5,8-Dibenzyloxy-6,7-dihydroxy-6,7-*O*-methylethylidene-2-(*N*-benzyl)-imino-1,3-diazonane (54). The guanidine 54 (139 mg, 94%) was obtained as a yellow foam from the thiourea 53 on a 287 μmol scale, according to the procedure described above for the guanidine 35. [α] -8 (c 0.5, CHCl₃); 1 H NMR: 1.26 (br.s, 6H, CH₃), 3.30–3.56 (m, 4H, H_{4,9}), 3.75 (m, 2H, NCH₂Ph), 4.07–4.58 (m, 8H, H_{5,6,7,8}, OCH₂Ph), 7.22 (br.s, 15H, CH_{Ph}); 13 C NMR: 26.8 (CH₃), 43.1, 46.0 (C_{4,9}, NCH₂Ph), 72.2 (OCH₂Ph), 79.0 (C_{5,6,7,8}), 109.8 (CMe₂), 128.0, 128.5 (CH_{Ph}), 137.3 (C_{Ph}); IR (neat): 1630 (v_{C=N}); HRMS (NH₃) for C₃₁H₃₈O₄N₃: (MH⁺) calcd 516.2862, found 516.2877.

(4*S*,5*R*,6*R*,7*S*)-4,7-Dibenzyloxymethyl-5,6-dihydroxy-5,6 -*O*-methylethylidene -2 - (*N*-butyl)-imino -1,3-diazepane (55). The guanidine 55 (318 mg, 65%) was obtained as a yellow foam from the thiourea 33 on a 1.02 mmol scale, according to the procedure described above for the guanidine 35 using *n*-butylamine. [α] + 42 (*c* 0.7, CHCl₃); 1 H NMR: 0.84 (m, 3H, H₄), 1.24–1.40 (m, 10H, H_{2',3'}, CH₃), 3.20–4.20 (m, 10H, H_{4,5,6,7,1'}, CH₂OBn), 4.19 (m, 4H, OCH₂Ph), 7.23 (m, 10H, CH_{Ph}); 13 C NMR: 13.6 (C_{4'}), 19.9 (C_{3'}), 27.1 (CH₃), 30.7 (C_{2'}), 41.0 (C_{1'}), 52.8 (C_{4,7}), 73.1 (CH₂OBn, OCH₂Ph), 77.5 (C_{5,6}), 109.9 (CMe₂), 127.6, 128.3 (CH_{Ph}), 138.0 (C_{Ph}), 159.0 (C₂); IR (neat): 1625 (ν_{C=N}); HRMS (NH₃) for C₂₈H₄₀O₄N₃: (MH⁺) calcd 482.3019, found 482.2986.

(4*S*,5*R*,6*R*,7*S*)-5,6-Dihydroxy-4,7-hydroxy methyl-5,6-*O* -methylethylidene-2-(*N*-butyl)-imino-1,3-diazepane (56). The guanidine 56 (140 mg, 73%) was obtained as an hygroscopic solid from the guanidine 55 on a 0.64 mmol scale, according to the procedure described above for the guanidino-sugar 36. [α] +45 (c 0.7, MeOH); ¹H NMR (CD₃OD): 0.93 (t, 3H, $J_{3',4'}$ =7.1 Hz, $H_{4'}$), 1.39–1.45 (m, 8H, $H_{3'}$, CH₃), 1,55 (m, 2H, $H_{2'}$), 3.14 (m, 2H, $H_{1'}$), 3.55–3.80 (m, 6H, $H_{4,7}$, CH₂OH), 4.05 (br.s, 2H, $H_{5.6}$); ¹³C NMR (CD₃OD): 16.2 (C_{4'}), 23.2 (C_{3'}), 29.3

(CH₃), 35.3 (C₂'), 45.9 (C₁'), 58.6 (C_{4,7}), 67.7 (CH₂OH), 80.9 (C_{5,6}), 112.1 (CMe₂), 159.5 (C₂); MS (NH₃): 302 (MH $^+$).

Methyl (4'S,5'R,6'R,7'S)-6-deoxy-6-(4',7'-dibenzyloxymethyl-5',6'-dihydroxy-5',6'-O-methylethylidene-1',3'-diazepan-2'-yl)-imino-2,3,4-tri-O-benzyl-D-glucopyranoside (57). The guanidine 57 (327 mg, 86%) was obtained as a yellow foam from the thiourea 33 on a 0.79 mmol scale, according to the procedure described above for the guanidine 35 using methyl 6-amino-6-deoxy-2,3,4-tri-O-benzyl-D-glucopyranoside. [α] + 36 (c 0.5, CHCl₃); ¹H NMR: 1.37 (m, 6H, CH₃), 3.25-4.10 (m, 17H, H_{4,5,6,7,2',3',4',5'}, CH₂N, OCH₃, CH₂OBn), 4.39-5.00 (m, 11H, OCH₂Ph, H₁'), 7.24 (m, 25H, CH_{Ph}); ¹³C NMR: 27.0 (CH₃), 46.0 (CH₂N), 54.0, 55.3 (C_{4,7}, OCH₃), 69.5 (C_{5,6}), 73.3, 74.9, 75.7 (CH₂OBn, OCH₂Ph), 77.3, 78.8, 80.1, 81.3 $(C_{2',3',4',5'})$, 98.0 $(C_{1'})$, 110.0 (CMe_2) , 127.7, 128.0, 128.4 (CH_{Ph}), 138.1, 138.7 (C_{Ph}), 158.0 (C₂); IR (neat): 1635 $(v_{C=N})$; HRMS (NH₃) for $C_{52}H_{62}O_9N_3$: (MH⁺) calcd 872.4486, found 872.4501.

Methyl (4'S,5'R,6'R,7'S)-6-deoxy-6-(4',7'-dihydroxymethyl-5',6'-dihydroxy-5',6'-O-methylethylidene-1',3'-diazepan-2'-yl)-imino-D-glucopyranoside (58). The guanidine 58 (209 mg, 79%) was obtained as an hygroscopic solid from the guanidine 57 on a 0.62 mmol scale according to the procedure described above for the guanidino-sugar 36. [α] +55 (c 0.75, CH₃OH); 1 H NMR (CD₃OD): 1.38 (s, 3H, CH₃), 1.46 (s, 3H, CH₃), 3.42 (m, 5H, CH₂N, OCH₃), 3.55–3.85 (m, 8H, H_{4,5,6,7}, CH₂OH), 3.90–4.40 (m, 4H, H_{2',3',4',5'}), 4.68 (m, 1H, H_{1'}); 13 C NMR (CD₃OD): 30.0 (CH₃), 47.0 (CH₂N), 51.8, 58.3 (C_{4,7}, OCH₃), 66.0 (CH₂OH), 76.1, 76.9, 77.4, 79.8, 81.7 (C_{5,6,2',3',4',5'}), 103.8 (C_{1'}), 112.0 (CMe₂), 159.9 (C₂).

Inhibition analysis of compounds 1, 2, 3, 4, 5 and 6 against various glycosidases. α-D-glucosidase from bacillus stearothermophilus (11×10^{-3} unit, EC 3.2.1.20), β -D-glucosidase from almonds (15×10^{-3} unit, EC 3.2.1.21), α -D-mannosidase from Jack beans (10×10^{-3} unit, EC 3.2.1.24) or α -L-fucosidase from bovine kidney (4 × 10⁻³ unit, EC 3.2.1.111) were purchased from Sigma-Aldrich Chimie. Incubation mixtures (1 mL) contained 0.05 M citrate-phosphate buffer at a pH of 6.8, 5.0, 4.5 and 5.5, respectively, according to the enzyme, 4-nitrophenyl-α-Dglucopyranoside (2 mM), or 4-nitrophenyl-β-D-glucopyranoside (2 mM), or 4-nitrophenyl-α-D-mannopyranoside (2 mM) or 4-nitrophenyl α -L-fucopyranoside (0.25 mM), the potential inhibitor 1, 2, 3, 4, 5, or 6 at a final concentration of 1 mM and finally, the enzyme α-D-glucosidase, 11×10^{-3} unit; β -D-glucosidase, 15×10^{-3} unit; α -D-mannosidase, 10×10^{-3} unit; or α -L-fucosidase, 4×10^{-3} unit per sample). After a 10 min incubation period at 37 °C, the reaction was quenched by the addition of a 0.2 M glycine-sodium hydroxide buffer at pH 10 (1 mL). The optical absorbance at 400 nm was measured to determine the amount of liberated 4-nitrophenol and the percentage of inhibition was calculated.

When the percentage of inhibition was greater than 50%, as it was the case for the compounds 5 and 6 against α -L-fucosidase, the assay was performed in presence of

variable amounts of the 4-nitrophenyl- α -L-fucopyranoside (in the range of 0.05 to 0.5 mM) and in the absence or in the presence of the inhibitor (at a constant final concentration of 1 mM) in 0.05 M citrate—phosphate buffer at pH 5.5. Lineweaver—Burk plots were then constructed for both reactions (with and without inhibitor) and the K_i was calculated from the two Michaelis—Menten constants (K_m) thus obtained.

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References and Notes

- 1. Kordik, C. P.; Reitz, A. B. J. Med. Chem. 1999, 42, 181.
- 2. Gross, P. E., Baker, M. A., Carver, J. P., Dennis, J. W. Clin. Cancer Res. 1995, 1, 935 and references cited.
- 3. van den Broek, L. A. G. M. In *Carbohydrates in Drug Design*; Witczak, Z. J., Nieforth, K. A., Eds; Dekker: New York, 1997.
- 4. Sinnott, M. L. Chem. Rev. 1990, 90, 1171.
- 5. Legler, G. Adv. Carbohydr. Chem. Biochem. 1990, 48, 319.
- 6. Winchester, B.; Fleet, G. W. J. Glycobiology 1992, 2, 199.
- 7. Jespersen, T. M.; Bols, M.; Sierks, M. R.; Skrydstrup, T. *Tetrahedron* **1994**, *50*, 13449.
- 8. Bols, M. Tetrahedron Lett. 1996, 37, 2097.
- 9. Hansen, A.; Tagmose, T. M.; Bols, M. *Tetrahedron* **1997**, *53*, 697.
- 10. Liu, K. K-C.; Kajimoto, T.; Chen, L.; Zhong, Z.; Ichikawa, Y.; Wong, C. H. J. Org. Chem. 1991, 56, 6280.
- 11. Bols, M. Acc. Chem. Res. 1998, 31, 1.
- 12. Wong, C.-H. Acc. Chem. Res. 1999, 32, 376.
- 13. Takebayashi, M.; Hiranuma, S.; Kanie, Y.; Kajimoto, T.; Kanie, O.; Wong, C.-H. *J. Org. Chem.* **1999**, *64*, 5280.
- 14. Look, G. C.; Fotsch, C. H.; Wong, C.-H. Acc. Chem. Res. 1993, 26, 182.
- 15. Tong, M. K.; Papandreou, G.; Ganem, B. J. Am. Chem. Soc. 1990, 112, 6137.
- 16. Ganem, B. Acc. Chem. Res. 1996, 29, 340.
- 17. Blériot, Y.; Genre-Grandpierre, A.; Tellier, C. *Tetrahedron Lett.* **1994**, *35*, 1867.
- 18. Knapp, S.; Choe, Y.-H.; Reilly, E. Tetrahedron Lett. 1993, 34, 4443.
- 19. Suzuki, K.; Fujii, T.; Sato, K.-I.; Hasimoto, H. Tetra-hedron Lett. 1996, 37, 5921.
- 20. Ganem, B.; Papandreou, G. J. Am. Chem. Soc. 1991, 113, 8984.
- 21. Papandreou, G.; Tong, M. K.; Ganem, B. J. Am. Chem. Soc. 1993, 115, 11682.
- 22. Hoos, R.; Vasella, A.; Rupitz, K.; Withers, S. G. Carbohydr. Res. 1997, 298, 291.
- 23. Fotsch, C. H.; Wong, C.-H. Tetrahedron Lett. 1994, 35, 3481.
- 24. Jeong, J.-H.; Murray, B. W.; Takayama, S.; Wong, C.-H. J. Am. Chem. Soc. 1996, 118, 4227.
- 25. Dong, W.; Jespersen, T.; Bols, M.; Skrydstrup, T.; Sierks, M. *Biochemistry* **1996**, *35*, 2788.
- 26. Blériot, Y.; Dintinger, T.; Guillo, N.; Tellier, C. *Tetrahedron Lett.* **1995**, *36*, 5175.

- 27. Blériot, Y.; Dintinger, T.; Genre-Grandpierre, A.; Padrines, M.; Tellier, C. *Bioorg. Med. Chem. Lett.* **1995**, *5*, 2655.
- 28. Lehmann, J.; Rob, B. Tetrahedron: Asymmetry 1994, 5, 2255
- 29. Kobayashi, Y.; Miyazaki, H.; Shiozaki, M. J. Am. Chem. Soc. 1992, 114, 10065.
- 30. Asano, N.; Kizu, H.; Oseki, K.; Tomioka, E.; Matsui, K.; Okamoto, M.; Baba, M. J. *Med. Chem.* **1995**, *38*, 2349.
- 31. Preliminary results: Gauzy, L., Le Merrer, Y., Depezay, J.C. Synlett 1998, 402.
- 32. Le Merrer, Y.; Poitout, L.; Depezay, J. C.; Dosbaa, I.; Geoffroy, S.; Foglietti, M. J. Bioorg. Med. Chem. 1997, 5, 519.
- 33. Nishimura, T.; Kitagima, K. J. Org. Chem. 1979, 44, 818.
- 34. Murphy, P. J.; Williams, H. L.; Hibbs, D. E.; Hursthouse, M. B.; Malik, K. M. A. *Tetrahedron* **1996**, *52*, 8315.
- 35. Hebrard, P.; Olomucki, M. *Bull. Soc. Chim. Fr.* **1970**, *5*, 1938. 36. Fritsche-Lang, W.; Wilharm, P.; Hädicke, E.; Fritz, H.; Prinzbach, H. *Chem. Ber.* **1985**, *118*, 2044.
- 37. Gravier-Pelletier, C., Bourissou, D., Le Merrer, Y., Depezay, J. C. Synlett 1996, 275.
- 38. Le Merrer, Y.; Duréault, A.; Gravier, C.; Languin, D.; Depezay, J-C. *Tetrahedron Lett.* **1985**, *26*, 319.
- 39. Le Merrer, Y.; Duréault, A.; Greck, C.; Micas-Languin, D.; Gravier, C.; Depezay, J.-C. *Heterocycles* **1987**, *25*, 541.
- 40. Greenhalgh, R.; Bannard, R. A. B. Can J. Chem. 1961, 39, 1017.
- 41. Dodd, D. S.; Kozikowski, A. P. Tetrahedron Lett. **1994**, 35, 977.
- 42. Chen, L.; Trilles, R. V.; Tilley, J. W. *Tetrahedron Lett.* **1995**, *36*, 8715.
- 43. Chittenden, J. F. G. Carbohydrate Res. 1980, 84, 350.
- 44. Sinclair, H. B. Carbohydrate Res. 1970, 12, 150.
- 45. Bagett, N., Stribbehill, P. J. Chem. Soc., Perkin Trans. I 1977, 1123.
- 46. David, S.; Hannessian, D. Tetrahedron 1985, 41, 643.
- 47. Garegg, P. J.; Lindberg, B. Carbohydrate Res. 1988, 173, 205.
- 48. Zhelvakova, E. G.; Magnashevskii, V. A.; Ermakova, L. I.; Shvets, V. I.; Preobrazhenskii, N. A. *J. Org. Chem. USSR* **1970**. *6*, 1997.
- 49. Zhelvakova, E. G.; Magnashevskii, V. A.; Ermakova, L. I.; Shvets, V. I.; Preobrazhenskii, N. A. *Zh. Org. Kh.* **1970**, *6*, 1987
- 50. Rossano, L. T.; Lo, Y. S.; Anzalone, L.; Lee, Y.-C.; Meloni, D. J.; Moore, J. R.; Gale, T. M.; Arnett, J. F. *Tetrahedron Lett.* **1995**, *36*, 4967.
- 51. Beck, A. K.; Bastani, B.; Plattner, D. A.; Petter, W.; Seebach, D.; Braunschweiger, H.; Gysi, P.; La Vecchia, L. *Chimia* **1991**, *45*, 238.
- 52. Allen, C. F. H.; Edens, C. O.; Vanallan, J. *Org. Syn. Coll.* **1955**, *III*, 394.
- 53. Schroeder, D.C. Chem. Rev. 1954, 181.
- 54. Bandelin, F. J.; Tuschhoff, J. V. J. Am. Chem. Soc. 1952, 74, 4271.
- 55. Finnegan, W. G.; Henrt, R. A.; Lieber, E. J. Org. Chem. **1953**, 18, 779.
- Lindel, T.; Hoffmann, H. Tetrahedron Lett. 1997, 38, 8935.
 Atwal, K. S.; Ahmed, S. Z.; O'Reilly, B. C. Tetrahedron Lett. 1989, 30, 7313.
- 58. Klamann, D.; Drahowzal, F. *Monatsh.* **1952**, *83*, 463 (*Chem. Abs.*) **1952**, *47*, 2707.
- 59. Shearer, B. G.; Lee, S.; Franzmann, K. W.; White, H. A. R.; Sanders, D. C. J.; Kiff, R. J.; Garvey, E. P.; Furfine, E. S. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 1763.
- 60. Astles, P. C.; Brown, T. J.; Cox, P.; Halley, F.; Lockey, P. M.; McCarthy, C.; McLay, I. M.; Majid, T. N.; Morley, A. D.; Porter, B.; Ratcliffe, A. J.; Walsh, R. J. A. *Bioorg. Med. Chem. Lett.* **1997**, *7*, 907.
- 61. Kim, K. S.; Qian, L. Tetrahedron Lett. 1993, 34, 7677.

- 62. Satoh, T.; Nishimura, Y.; Kondo, S.; Takeuchi, T. J. Antibiot. 1995, 45, 321.
- 63. Levallet, C.; Lerpinière, J.; Ko, S. Y. *Tetrahedron* **1997**, *53*, 5291.
- 64. Avalos, M.; Babiano, R.; Cintas, P.; Duran, C. J.; Jimenez, J. L.; Palacios, J. C. *Tetrahedron* 1995, 51, 8043.
- 65. Whistler, R. L.; Wolfrom, M. L. Methods in Carbohydrate Chemistry 1963, 2, 244.
- 66. To facilitate the notation in the attribution of ${}^{1}H$ NMR signals of C_2 symmetric molecules, the coupling constants are reported only for the half of the molecule while all the protons

are attributed. For example, in the molecule **14**, H_4 and H_9 have the same chemical shift as well as $H_{4'}$ and $H_{9'}$. The corresponding notation for these protons is: 3.42 (dd, 2H, $J_{4,5} = 5.8$, $J_{4,4'} = 15.6$ Hz, $H_{4,9}$), 3.64 (m, 2H, $H_{4',9'}$).

67. Charette, A. B.; Côté, B. J. Org. Chem. 1993, 58, 933.